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## **STRATIGRAPHIC CONSULTANTS**

### **THE TROLL FIELD**

**Analysis of biostratigraphic data by  
RASC and STRATCOR**

**Summary**

**Ron Woollam**

# SUMMARY

- OBJECTIVE

- review Jurassic and Lower Cretaceous palynostratigraphy of the Troll Field
- achieved through application of quantitative techniques

- DATABASE

- 28 wells / 5000 samples
- 186 bioevents / 1479 event records

- JGR SCHEME

- routine stratigraphic subdivision of wells
- inconsistent correlation framework

- PC BASED RASC & STRATCOR

- statistical and graphical techniques
- testing of JGR scheme (RASC)
- consistent high resolution correlation framework (STRATCOR)

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## **INTRODUCTION**

The project was undertaken on behalf of A/S Norske Shell. The overall objective was to critically review palynological data from a total of twenty-eight wells in the Troll Field. The interval studied covers the Jurassic to lowermost Cretaceous. Data analysis was undertaken by computer-based quantitative methods (RASC and STRACOR).

The results of previous biostratigraphic work are embodied in the Joint Geological Review biozonation. The JGR scheme was developed from earlier zonations, through the collaborative efforts of the Troll Field operating companies, after almost a decade of independent work. The zonation comprises thirty-five events based on dinocyst inceptions, extinctions and abundance changes. A total of ten zones/subzones are recognized. While the JGR scheme allows routine subdivision of individual well sections, many inconsistencies exist when correlating from one section to another.

## **RASC**

Statistical analysis using RASC highlights the limited geographic continuity of many of the bioevents as shown by the cumulative frequency curve and occurrence matrix. This situation is the norm rather than the exception - a characteristic of the fossil record. Despite the inconsistencies in correlating between sections, the succession of bioevents in the JGR scheme compares favourably with the optimum sequence derived by the RASC program.

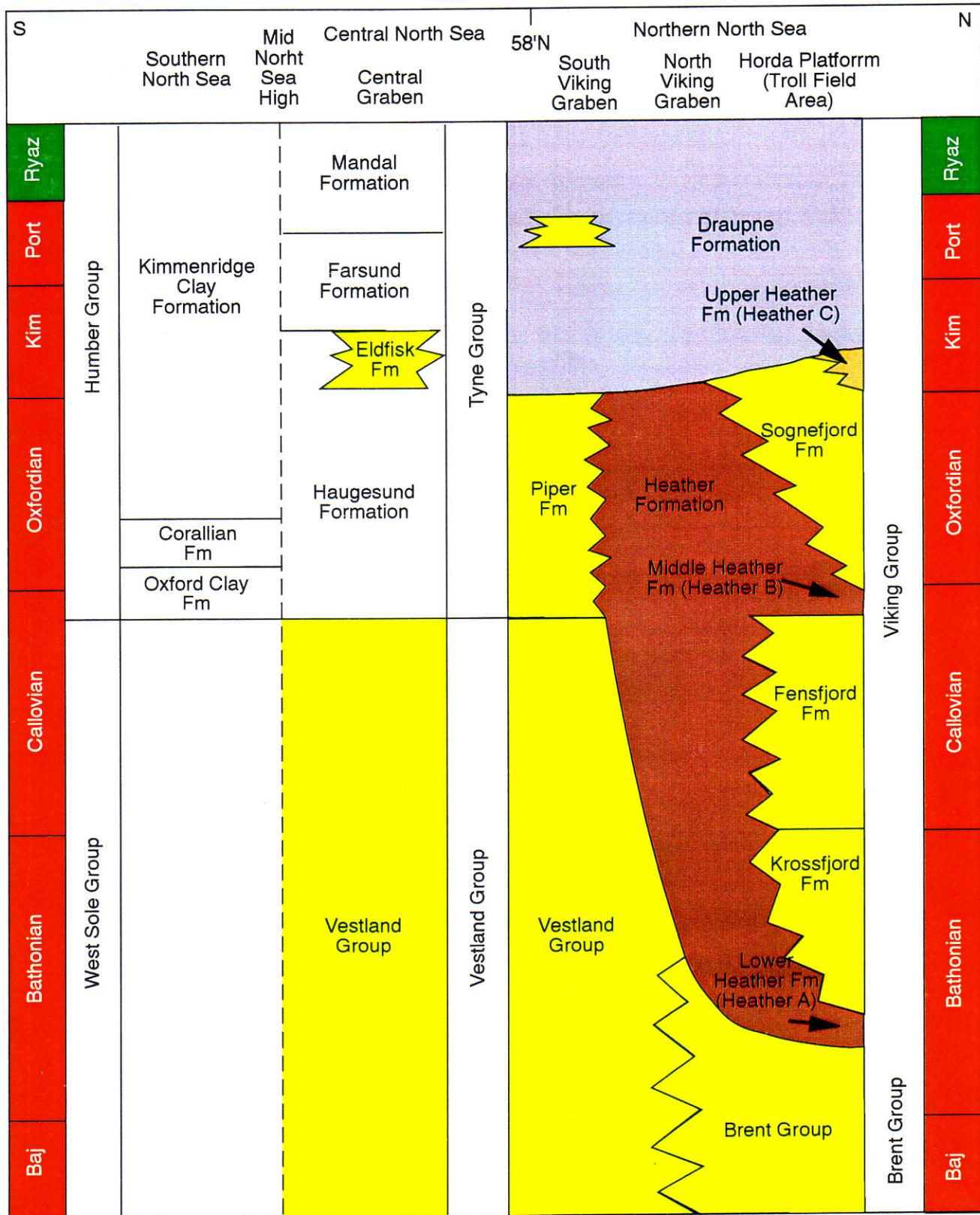
## **STRATCOR**

STRATCOR builds a biozonation by graphically compositing all wells in the database. The final composite standard sequence of bioevents is a probabilistic zonation similar to the RASC optimum sequence. Graphic correlation of the FCSS with each individual well, allows the zonation to be interpolated throughout the well database. The result is a high-resolution correlation framework. Plotting the deviations of observed from interpolated positions for each bioevent reveals a normal distribution of values, and illustrates the probabilistic nature of the technique. Reversing the correlation procedure from the individual well sections to the composite also enables data to be collected on the distributions of individual events, scaled in FCSS units.

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# LITHOSTRATIGRAPHY

After JGR (1988) report with minor modifications



Note: Thicknesses are schematic

# PREVIOUS BIOZONATIONS

- NORSKE SHELL SCHEME (1983)
- PL085 SCHEME (1984)
- AMALGAMATED INTO JGR SCHEME (1988)
  - 35 bioevents (dinocyst inceptions, extinctions and abundance changes)
  - 7 zones / 6 subzones = 10 divisions
  - 17 correlation lines (biohorizons)
- SUBSET OF TOTAL DATABASE
- BASIC SUBDIVISION OF STRATIGRAPHY
- INCONSISTENT CORRELATION FRAMEWORK (DUE TO FOSSIL RECORD - NOT ANALYSIS)
- PROBLEMS OF PICKING BIOZONES IN INDIVIDUAL WELLS DUE TO "OUT OF SEQUENCE" BIOEVENTS
- SUBJECTIVE BIOZONE CORRELATION

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# JGR BIOZONATION

After JGR report with minor modifications and taxonomic update

Ma (Harland et al., 1982)	AGE	TROLL AREA ZONES AND SUBZONES	VALIDITY	DETECTABILITY	RELIABILITY (*)	BOUNDARY CRITERIA	PL085	LITHO STRATIGRAPHY		FORMATION		
								SHELL	ZONE			
150 [140]	EARLY VOLGAN	KIMMERIDGIAN ( <i>sensu gallico</i> )	<i>Oligosphaeridium pulcherrimum</i> s.l.	1	2	3	FDA <i>Oligosphaeridium patulum</i> (= <i>O. pulcherrimum</i> s.l.)	T8	9.3	7C 7M	HEATHER FORMATION UNIT-C	
				3	3	6	FDA <i>Leptodinium subtile</i>					
			(Pu)	1	2	3	LDA <i>Egmontod. polyplacophorum</i> (reg.) LDA <i>Oligosphaeridium patulum</i>					
			<i>Leptodinium subtile</i>	2	3	5	FDA <i>Leptodinium subtile</i> (reg.) FDA <i>Sentusidinium pilosum</i> (reg.)	T7	9.2	7 6 6M		
				3	3	6	LDA <i>Hystrichodinium pulchrum</i> LDA <i>Cyclonephelium distinctum</i> Group					
			<i>Scriniodinium crystallinum</i> (Cr)	2	2	4	FDA <i>Scriniodinium crystallinum</i> LDA <i>Ambonosp. cf. calloviana</i> (acme) LDA <i>Criboperidinium granulatum</i> (reg.)	T6	8.3 8.2	6 6C 6M		
				2	1	3	FDA <i>Endoscrinium galeritum</i> FDA <i>Lithodinia</i> sp. B					
			<i>Endoscrinium galeritum</i> (Ga)	3	3	6	FDA <i>Glossodin. aff. dimorphum</i> (local) FDA <i>Dinocyst</i> sp. A (local) FDA <i>Endoscrinium galeritum</i> (common) LDA <i>Dinocyst</i> sp. A (local) LDA <i>Glossodin. aff. dimorphum</i> (local)			5 5M		
				3	2	5	LDA <i>Occisucysta cf. evittii</i> LDA <i>Systematophora vestitum</i> (reg.)					
156 [145]	LATE OXFORDIAN		<i>Adnatosphaeridium aenulum</i>	2	2	4	FDA <i>Rigaudella aemula</i> (= <i>A. aemulum</i> ) FDA <i>Compositosphaeridium polonicum</i>	T5	8.1	4 4M	SOGNEFJORD FM.	
				3	2	5	LDA <i>Occisucysta cf. evittii</i> LDA <i>Systematophora vestitum</i> (reg.)					
			<i>G. jurassica</i> var. <i>longicornis</i> (AeL)	2	1	3	FDA <i>G. jurassica</i> var. <i>longicornis</i> LDA <i>Leptodinium mirabile/subtile</i> Group LDA <i>Lithodinia</i> sp. B LDA <i>G. jurassica</i> var. <i>longicornis</i>	T4	7	4 4M		
				2	1	3	LDA <i>Leptodinium mirabile/subtile</i> Group LDA <i>Lithodinia</i> sp. B LDA <i>G. jurassica</i> var. <i>longicornis</i>					
			Acanthaulax senta	1	1	2	FDA <i>Liesbergia scarburghensis</i> (= <i>A.s.</i> ) FDA <i>Wanaea fimbriata</i>	T3	6.2 6.1	3 3M		
				3	2	5	LDA <i>R. aemula</i> (a.) / LDA <i>C. cf. cerastes</i>					
163 [152]	CALLOVIAN		<i>Systematophora divarica</i> (SeD)	3	2	5	FDA <i>Systematophora divarica</i> LDA <i>Wanaea fimbriata</i> LDA <i>Liesbergia scarburghensis</i>			HEATHER FORMATION UNIT B		
				1	1	2	LDA <i>Liesbergia scarburghensis</i>					
			<i>Lithodinia jurassica</i> (Ju)	1	2	3	FDA <i>Lithodinia jurassica</i>	T2	5	2 2M		
									1		FENSFJORD FM.	

Haq et al., (1987) ages in parentheses

REGIONAL BIOEVENT QUALITY			
VALIDITY (V)	1=High	2=Medium	3=Low
DETECTABILITY (D)	1=Good	2=Fair	3=Poor
RELIABILITY (V+D)	2/3=High	4=Medium	5/6=Low

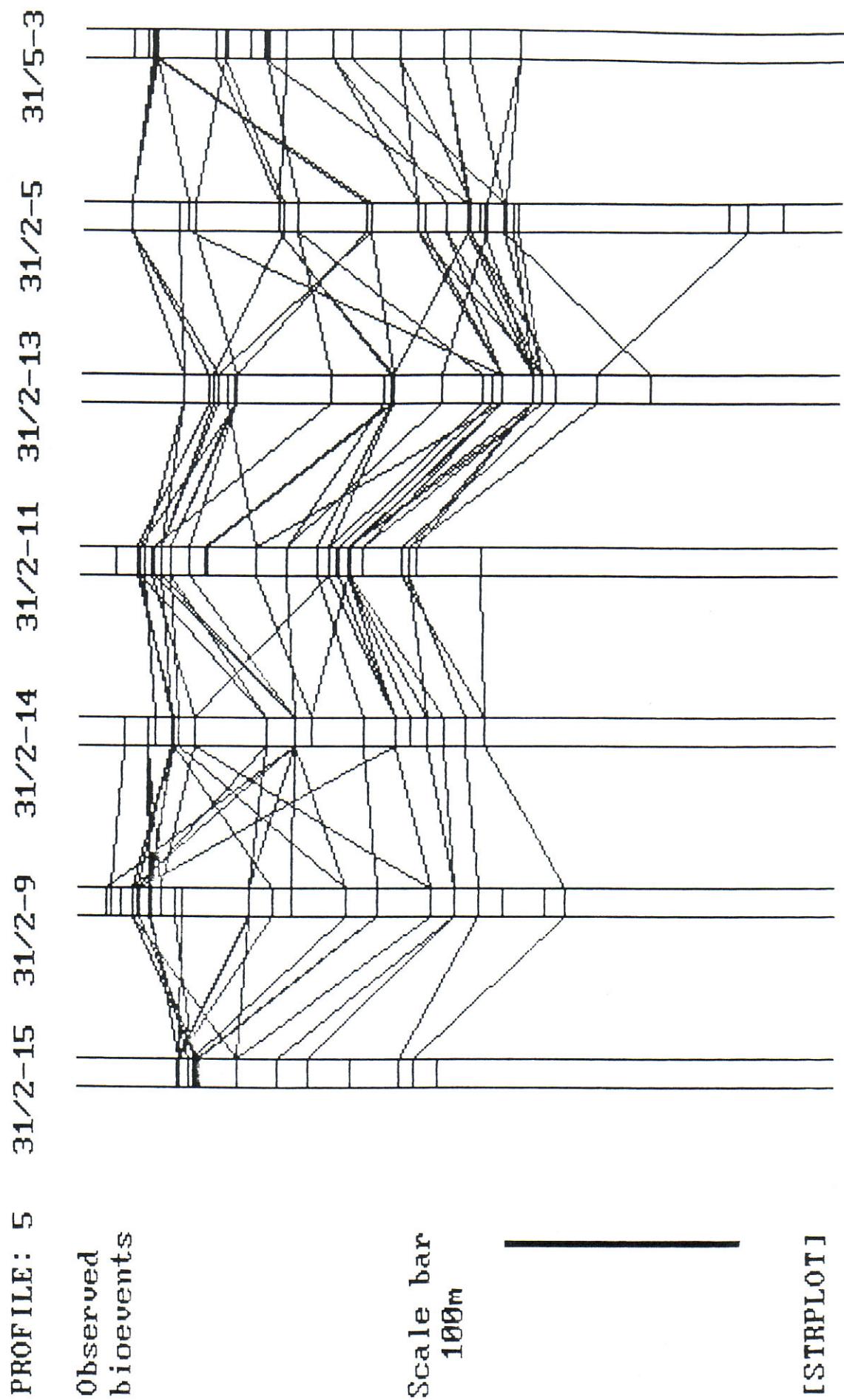
## **DATABASE EVALUATION**

- 28 WELLS
- LIST EVENTS
- 186 FDA/LDA BIOEVENTS (50% IN INTERVAL OF INTEREST)
- 1479 BIOEVENT RECORDS
- c. 5000 PALynological SAMPLES (CC, SWC)
- c. 2-4m SAMPLE INTERVALS
- NUMEROUS ANALYSTS

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[STRPLOT]

Scale bar  
100m





# **QUANTITATIVE METHODS**

## **PC APPLICATIONS**

- **RASC (RANKING & SCALING)**
  - statistical method
  - matrix transformations (ordering rows/columns) of cross-over frequencies for event pairs
  - scaling based on standardized normal distribution
  - scaled optimum sequence of events
  - 10 or more wells
  - short run time (RASC12 batch version)
- **STRATCOR (STRATIGRAPHIC CORRELATION)**
  - graphical interpolation method
  - composites well sections
  - final composite standard sequence
  - FCSS correlation in all wells
  - 2 or more wells
  - requires user input throughout (Version 1.6)
- **BIOZONATIONS SHOWN AS DENDROGRAMS**

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# RASC

- INPUT DATA

- event dictionary
- event sequence file
- depth not considered

- BIOEVENT DISTRIBUTION

- cumulative frequency of events
- event occurrence table

- SCALED OPTIMUM SEQUENCE (AVERAGE)

- test JGR sequence (maximum)
- average sequence = maximum sequence for robust, low variance bioevents
- also true for widely spaced bioevents

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**CUMULATIVE FREQUENCY DISTRIBUTION OF  
JURASSIC BIOSTRATIGRAPHIC  
EVENTS IN THE TROLL FIELD AREA (MAINLY DINOCYSTS)**

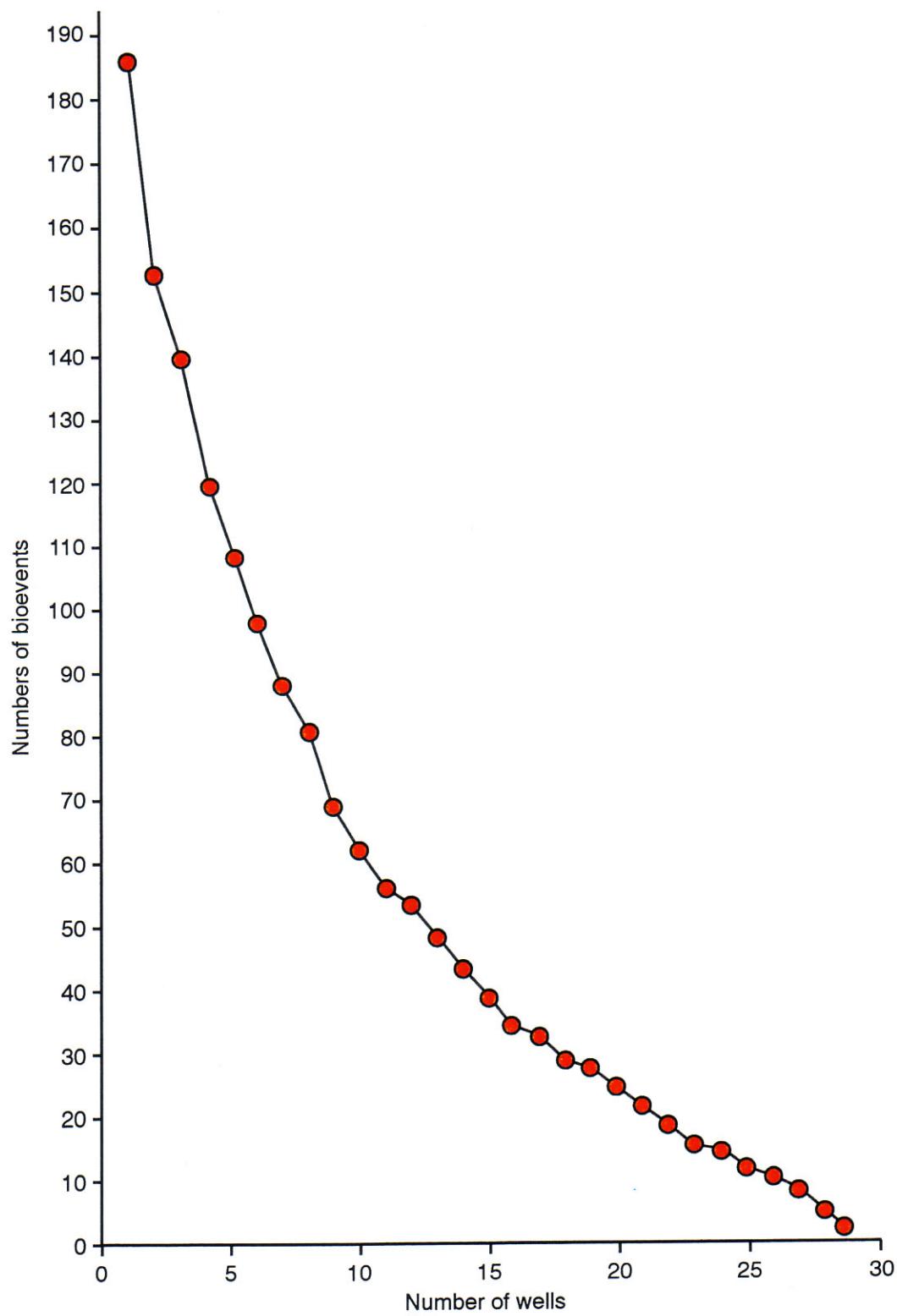
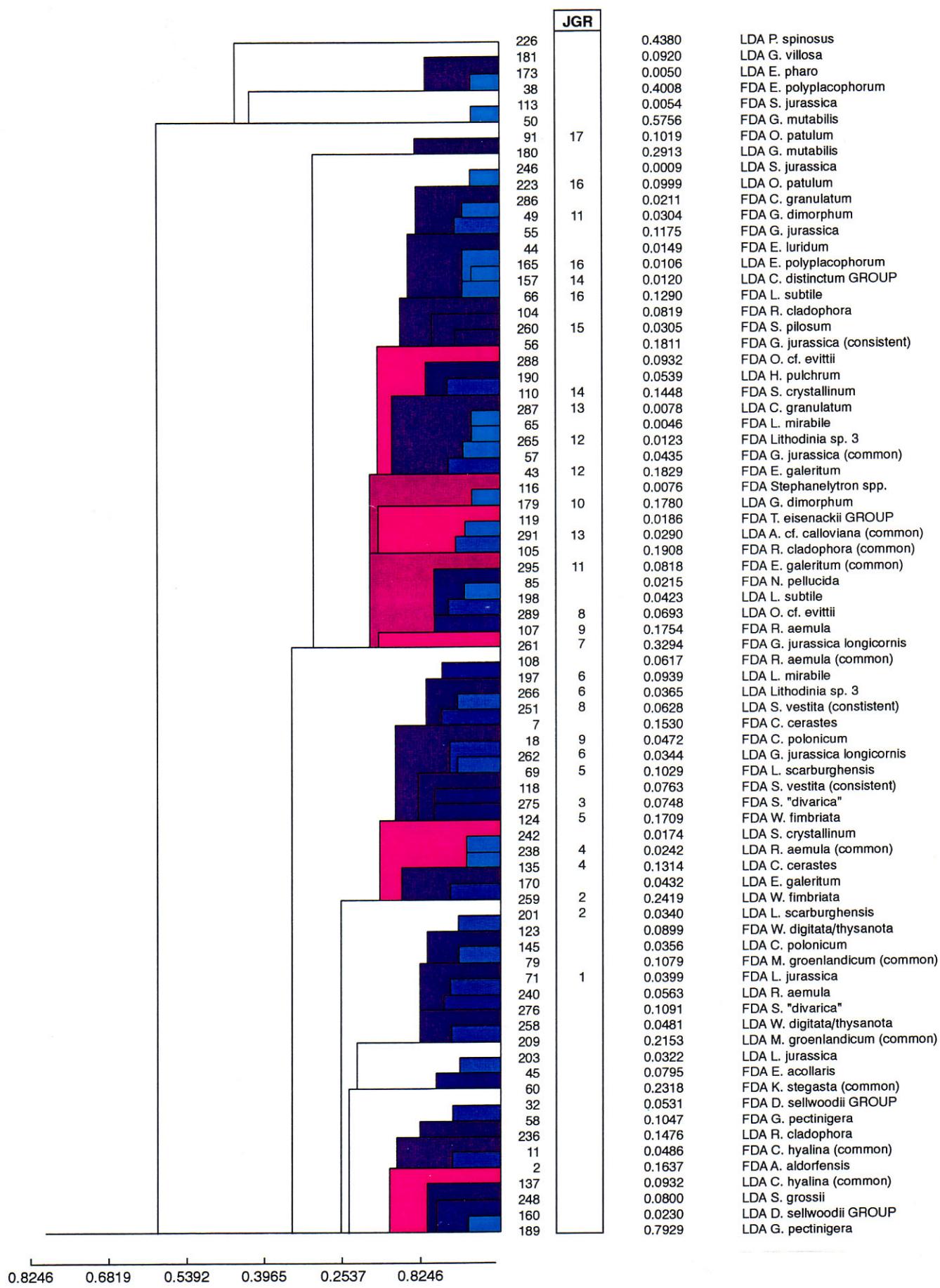
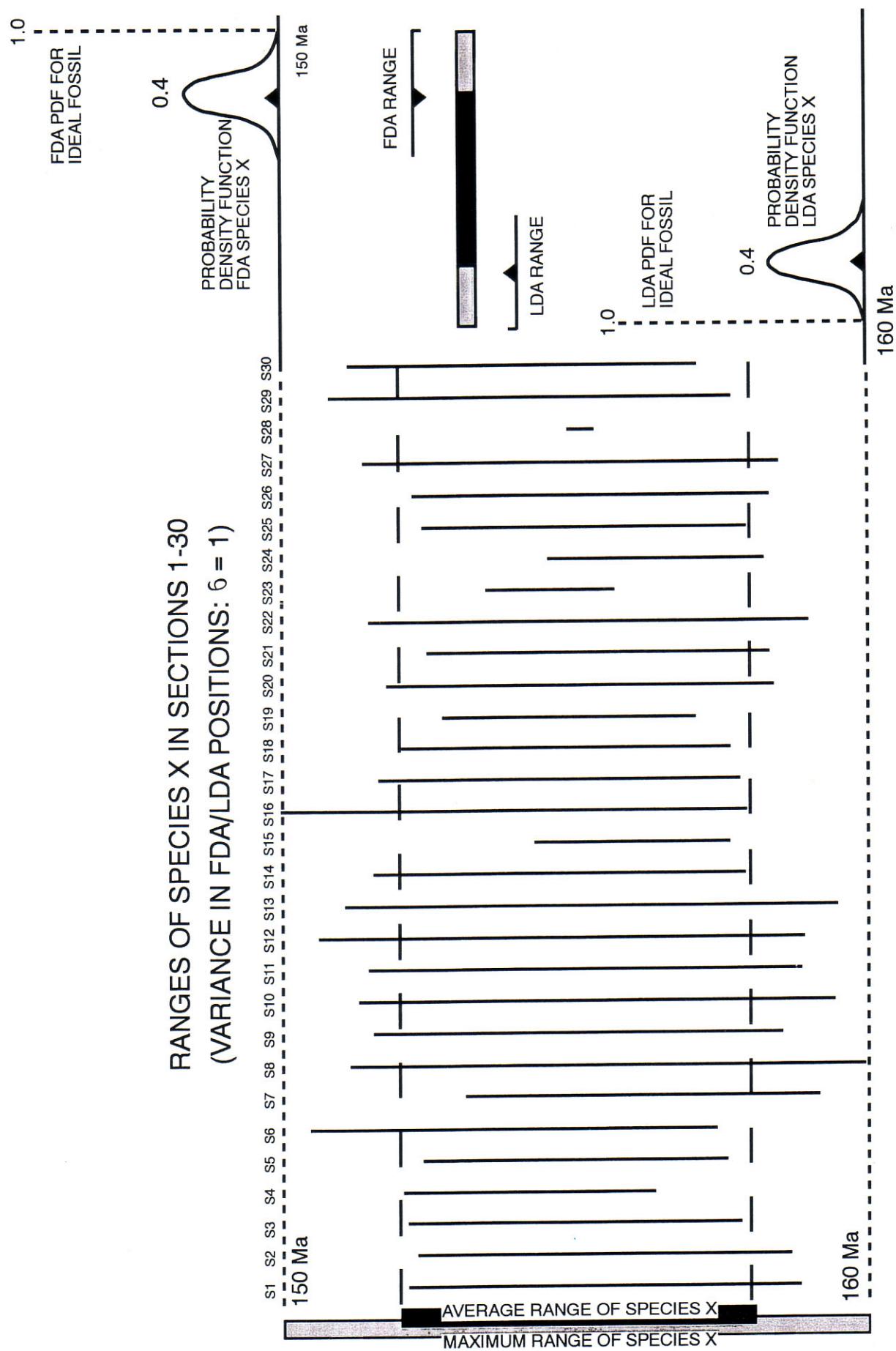


TABLE 4.1 EVENT OCCURRENCE MATRIX (RASC 8/1/4)

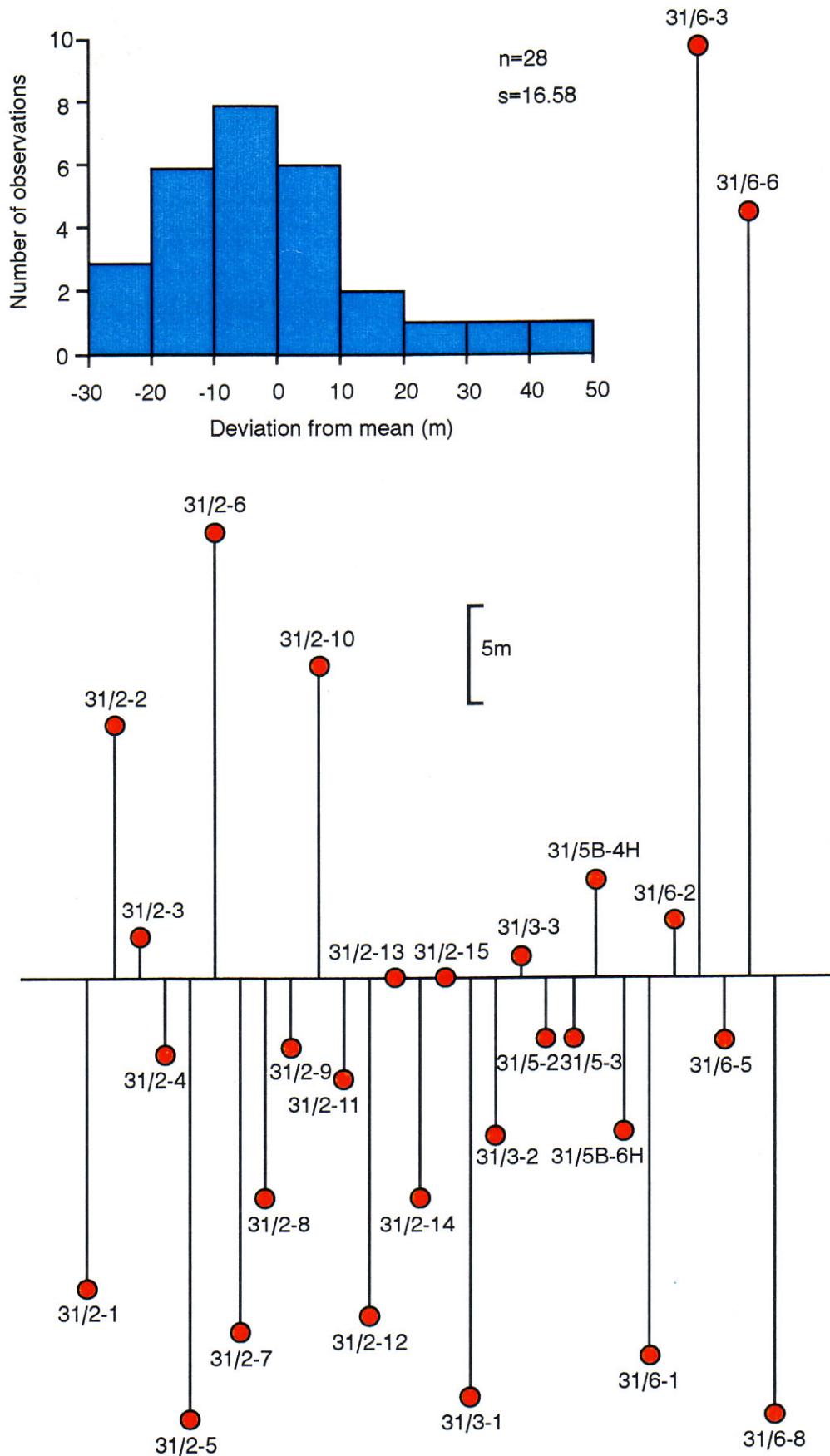
# DENDROGRAM OF WEIGHTED INTERFOSSIL DISTANCE (RASC 8/1/4)



# AVERAGE VERSUS MAXIMUM BIOEVENT RANGE



# DEVIATIONS OF OBSERVED EVENT POSITIONS FROM MEAN POSITION FOR FDA S. CRYSTALLINUM



# **STRATCOR**

## ● **INPUT DATA**

- event dictionary
- event sequence file
- event depth file

## ● **BIOZONATION (FCSS)**

- average bioevent positions
- 15 event clusters (biozones)
- potential for subdivision
- tie to sequences

## ● **CORRELATION**

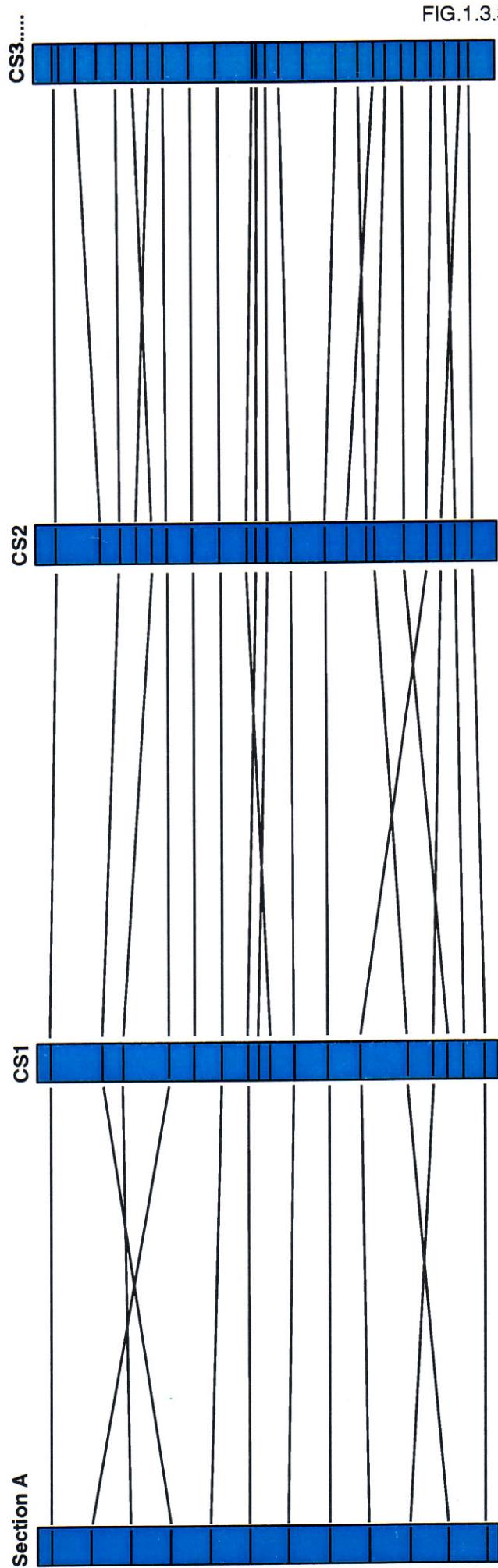
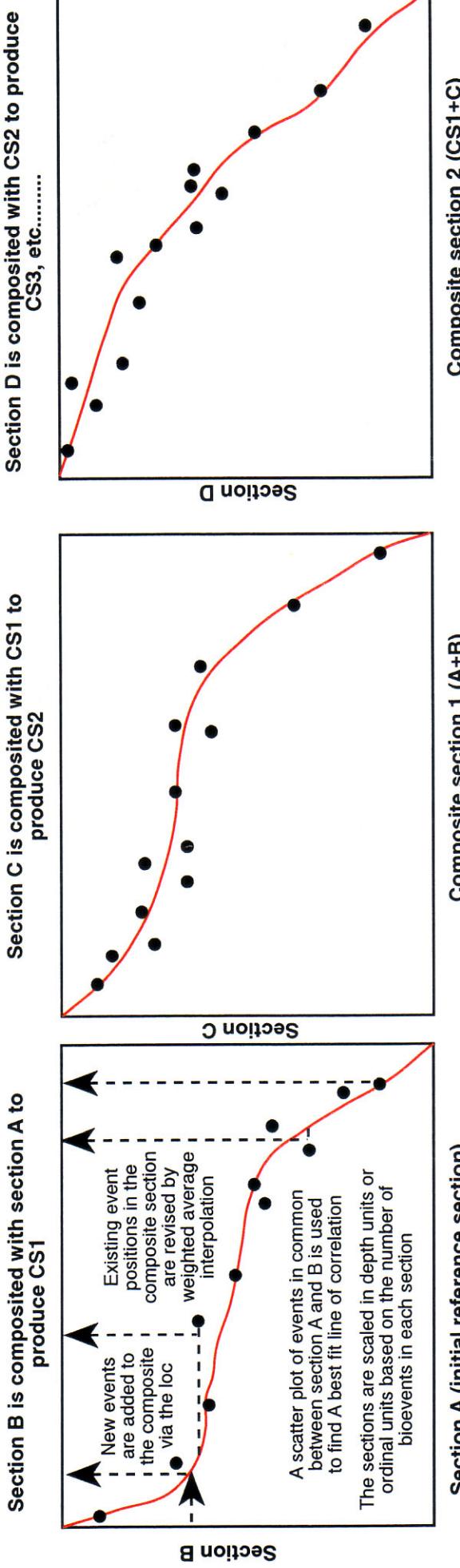
- 3000 interpolated event positions (over 2 x observed records)
- c. 120 events per well
- high resolution framework

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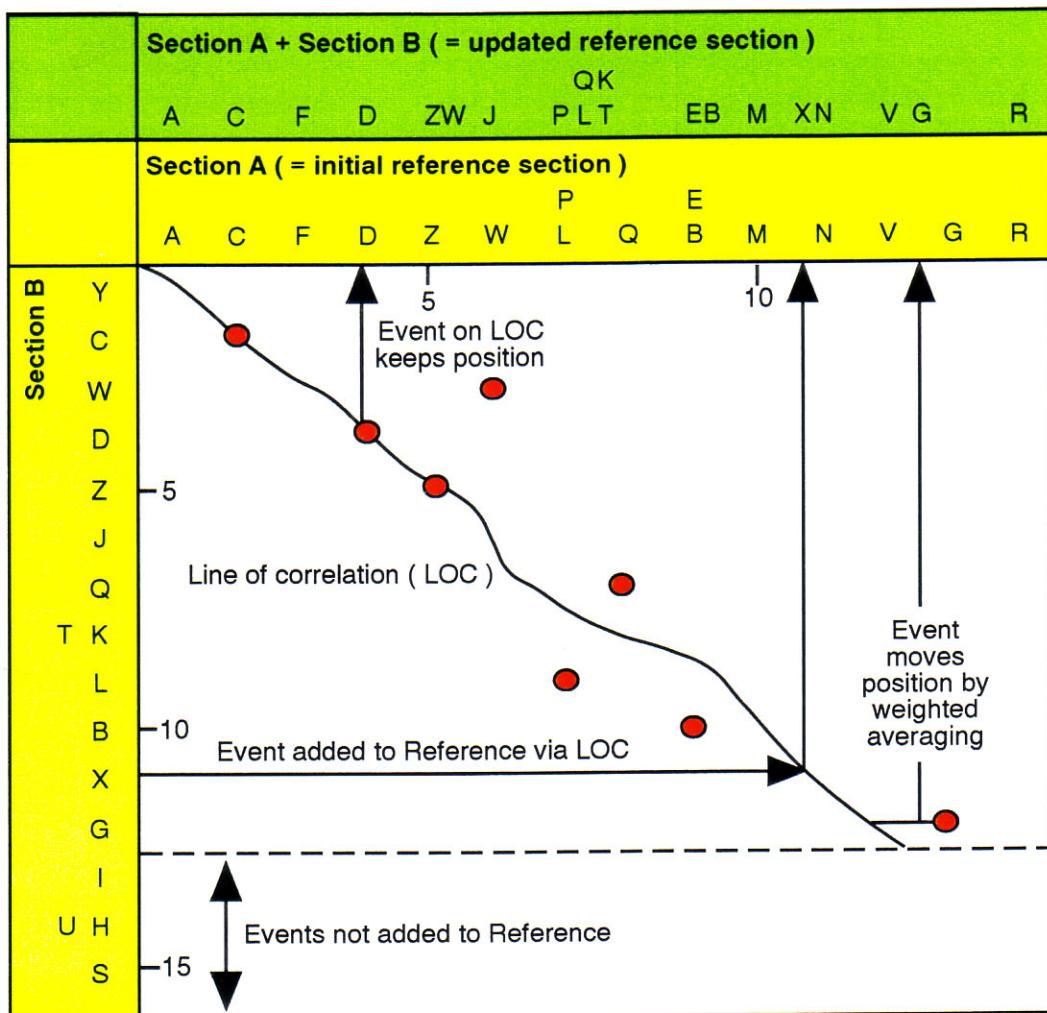
# PROBABILISTIC BIOZONATION (STRATCOR)

NSEP 93-15

FIG.1.3.3.3



# INTERPOLATION METHOD IN GRAPHIC CORRELATION (STRATCOR)

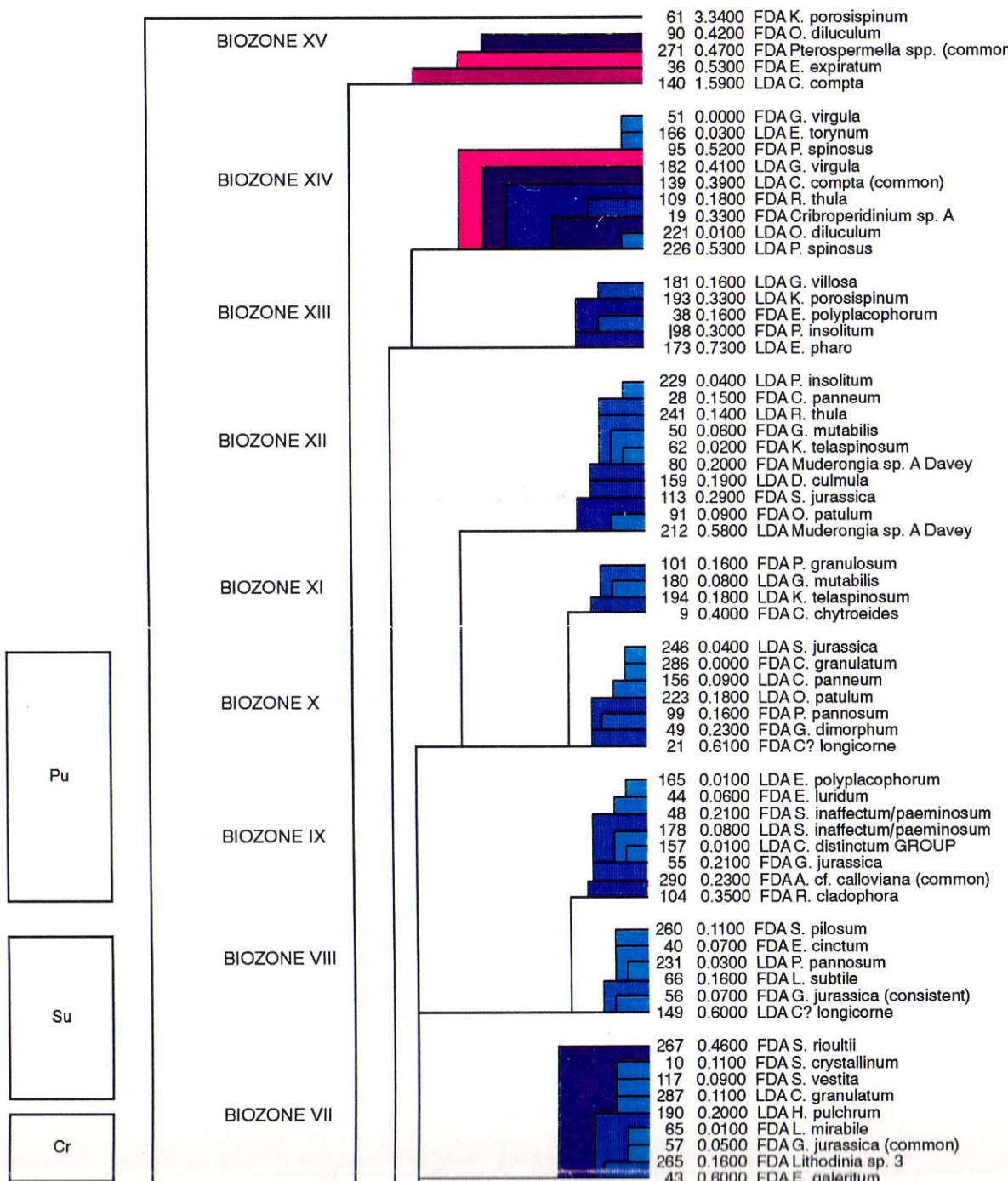


Initial Reference Section

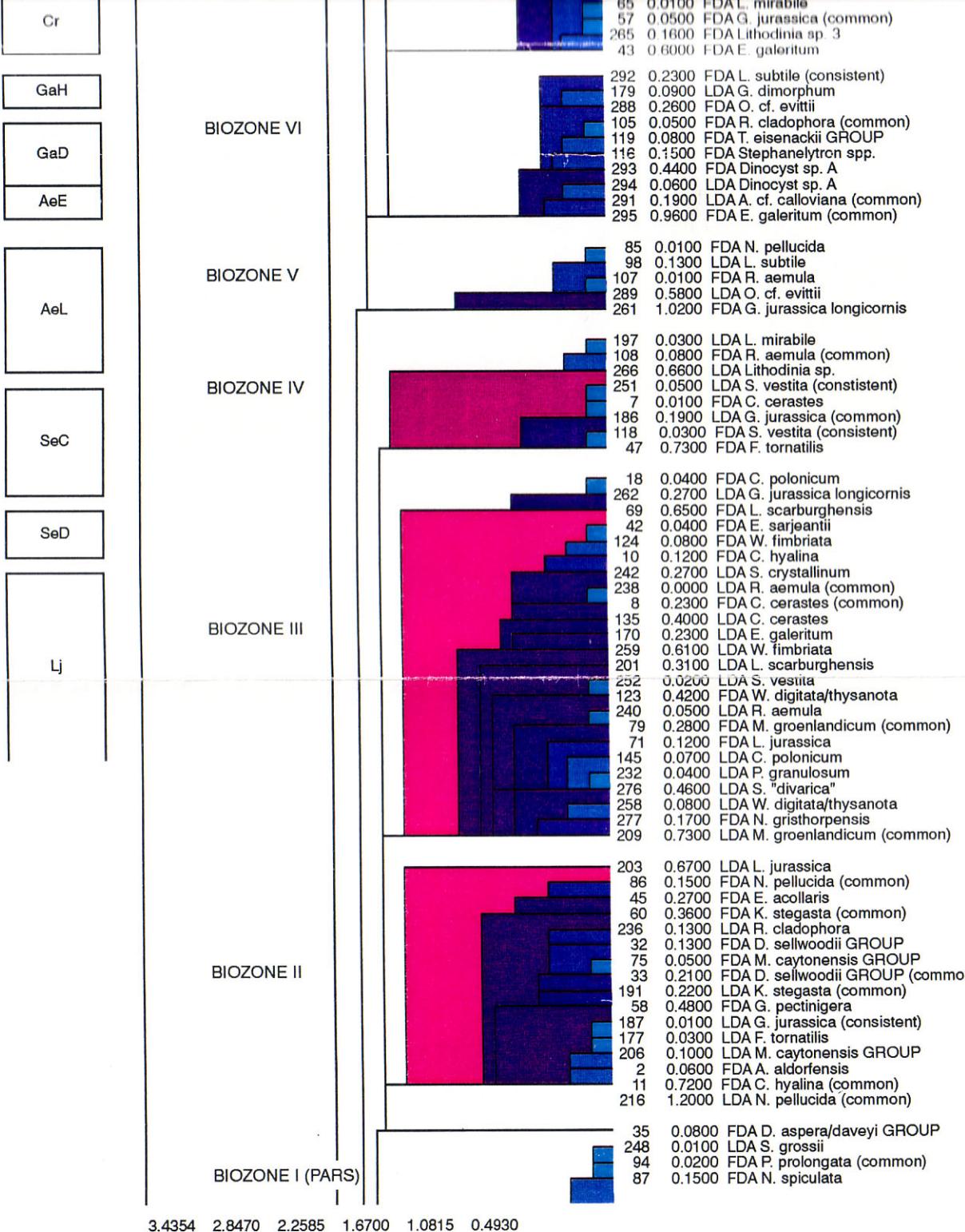
Event	Position
A	1.00
C	2.00
F	3.00
D	4.00
Z	5.00
W	6.00
L	7.00
P	7.00
Q	8.00
B	9.00
E	9.00
M	10.00
N	11.00
V	12.00
G	13.00
R	14.00

Updated Reference Section

Event	Position
A	1.00
C	2.00
F	3.00
D	4.00
Z	5.00
W	5.50
J	5.75
P	7.00
L	7.50
Q	7.50
T	7.75
K	7.75
E	9.00
B	9.25
M	10.00
X	10.75
N	11.00
V	12.00
G	12.75
R	14.00



## DENDROGRAM OF STRATCOR FCSS DIC DIST BIOEVENT (FCSSS)



## BIOEVENT SEQUENCE COMPARISON RASC - STRATCOR

### RASC OPTIMUM SEQUENCE

FDA *O. patulum*  
 LDA *O. patulum*  
 FDA *G. dimorphum*  
 LDA *E. polyplacophorum*  
 LDA *C. distinctum* Group  
 FDA *L. subtile*  
 FDA *S. pilosum*  
 LDA *H. pulchrum*  
 FDA *S. crystallinum*  
  
 LDA *C. granulatum*  
 FDA *Lithodinia sp. B*  
 FDA *E. galeritum*  
 LDA *G. dimorphum*  
 LDA *A. cf. calloviana* (c)  
  
 FDA *E. galentum* (c)  
 LDA *O. cf. evittii*  
 FDA *R. aemula*  
 FDA *G. j var. longicornis*  
 LDA *L. mirabile/subtile* Group  
 LDA *Lithodinia sp. B*  
 LDA *S. vestitum* (consistent)  
 FDA *C. polonicum*  
 LDA *G. j. var. longicornis*  
 FDA *L. scarburghensis*  
 FDA *S. divarica*  
 FDA *W. fimbriata*  
 LDA *R. aemula* (c)  
 LDA *C. cerastes*  
 LDA *W. fimbriata*  
 LDA *L. scarburghensis*  
 FDA *L. jurassica*

### STRATCOR FCSS

FDA *O. patulum*  
 LDA *O patulum*  
 FDA *G. dimorphum*  
 LDA *E. polyplacophorum*  
 LDA *C. distinctum* Group  
 FDA *S. pilosum*  
 FDA *L. subtile*  
 LDA *A. cf. calloviana* (c)  
 FDA *S. crystallinum*  
 LDA *C. granulatum*  
 LDA *H. pulchrum*  
 FDA *Lithodinia sp. B*  
 FDA *E. galeritum*  
 LDA *G. dimorphum*  
 FDA *Dinocyst sp. A*  
 Lda *Dinocyst sp. A*  
 FDA *E. galeritum* (c)  
 FDA *R. aemula*  
 LDA *O. cf. evittii*  
 FDA *G. j. var. longicornis*  
 LDA *L. mirabile/subtile* Group  
 LDA *Lithodinia sp. B*  
 LDA *S. vestitum* (consistent)  
 FDA *C. polonicum*  
 LDA *G. j. var. longcornis*  
 FDA *L. scarburghensis*  
 FDA *W. fimbriata*  
 FDA *S. divarica*  
 LDA *R. aemula* (c)  
 LDA *C. cerastes*  
 LDA *W. fimbriata*  
 LDA *L. scarburghensis*  
 FDA *L. jurassica*

# BIOEVENT SEQUENCE COMPARISON

## RASC-JGR-STRACTOR

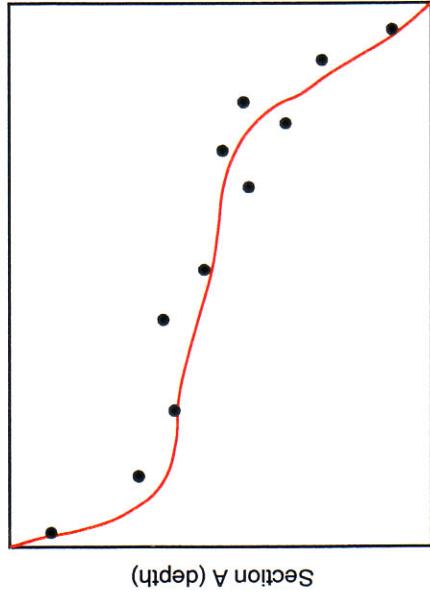
RASC OPTIMUM SEQUENCE	JGR BIOEVENTS	STRATCOR FCSS
FDA <i>O. patulum</i>	FDA <i>O. patulum</i>	FDA <i>O. patulum</i>
LDA <i>O. patulum</i>	FDA <i>L. subtile</i>	LDA <i>O. patulum</i>
FDA <i>G. dimorphum</i>	LDA <i>E. polyplacophorum</i> (reg.)	FDA <i>G. dimorphum</i>
LDA <i>E. polyplacophorum</i>	LDA <i>O. patulum</i>	LDA <i>E. polyplacophorum</i>
LDA <i>C. distinctum</i> Group	FDA <i>L. subtile</i> (reg.)	LDA <i>C. distinctum</i> Group
FDA <i>L. subtile</i>	FDA <i>S. pilosum</i> (reg.)	FDA <i>S. pilosum</i>
FDA <i>S. pilosum</i>	LDA <i>H. pulchrum</i>	FDA <i>L. subtile</i>
LDA <i>H. pulchrum</i>	LDA <i>C. distinctum</i> Group	LDA <i>A. cf. calloviana</i> (c)
FDA <i>S. crystallinum</i>	FDA <i>S. crystallinum</i>	FDA <i>S. crystallinum</i>
LDA <i>C. granulatum</i>	LDA <i>A. cf. calloviana</i> (acme)	LDA <i>C. granulatum</i>
FDA <i>Lithodinia sp. B</i>	LDA <i>C. granulatum</i> (reg.)	FDA <i>Lithodinia sp. B</i>
FDA <i>E. galeritum</i>	FDA <i>L. Lithodinia sp. B</i>	FDA <i>E. galeritum</i>
LDA <i>G. dimorphum</i>	FDA <i>E. galeritum</i>	LDA <i>G. dimorphum</i>
LDA <i>A. cf. calloviana</i> (c)	FDA <i>G. aff. dimorphum</i> (local)	FDA <i>Dinocyst sp. A</i>
FDA <i>E. galeritum</i> (c)	FDA <i>Dinocyst sp. A</i> (local)	LDA <i>Dinocyst sp. A</i>
	FDA <i>E. galeritum</i> (common)	FDA <i>E. galeritum</i> (c)
LDA <i>O. cf. evittii</i>	LDA <i>Dinocyst sp. A</i> (local)	
FDA <i>R. aemula</i>	FDA <i>R. aemula</i>	FDA <i>R. aemula</i>
FDA <i>G. j var. longicornis</i>	FDA <i>C. polonicum</i>	LDA <i>O. cf. evittii</i>
LDA <i>L. mirabile/subtile</i> Group	LDA <i>O. cf. evettii</i>	FDA <i>G. j. var. longicornis</i>
LDA <i>Lithodinia sp. B</i>	LDA <i>S. vestitum</i> (reg.)	LDA <i>L. mirabile/subtile</i> Group
LDA <i>S. vestitum</i> (consistent)	FDA <i>G. j. var. longicornis</i>	LDA <i>Lithodinia sp. B</i>
FDA <i>C. polonicum</i>	LDA <i>L. mirabile/subtile</i> Group	LDA <i>S. vestitum</i> (consistent)
LDA <i>G. j. var. longicornis</i>	LDA <i>Lithodinia sp. B</i>	FDA <i>C. polonicum</i>
FDA <i>L. scarburghensis</i>	LDA <i>G. j. var. longicornis</i>	LDA <i>G. j. var. longcornis</i>
FDA <i>S. divarica</i>	FDA <i>L. scarburghensis</i>	FDA <i>L. scarburghensis</i>
FDA <i>W. fimbriata</i>	FDA <i>W. fimbriata</i>	FDA <i>W. fimbriata</i>
LDA <i>R. aemula</i> (c)	LDA <i>R. aemula</i> (a.)	FDA <i>S. divarica</i>
LDA <i>C. cerastes</i>	LDA <i>C. cf. cerastes</i>	LDA <i>R. aemula</i> (c)
LDA <i>W. fimbriata</i>	FDA <i>S. divarica</i>	LDA <i>C. cerastes</i>
LDA <i>L. scarburghensis</i>	LDA <i>W. fimbriata</i>	LDA <i>W. fimbriata</i>
FDA <i>L. jurassica</i>	LDA <i>L. scarburghensis</i>	LDA <i>L. scarburghensis</i>
	FDA <i>L. jurassica</i>	FDA <i>L. jurassica</i>

# AGE CALIBRATION OF STRATCOR FCSS

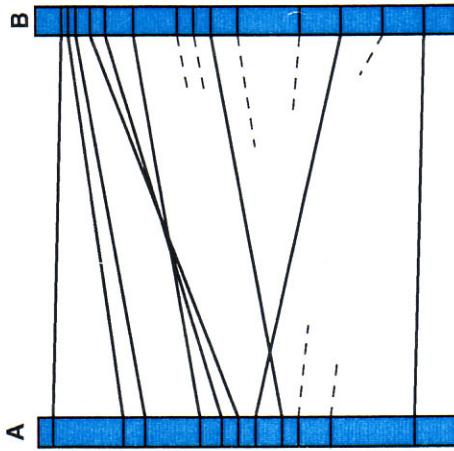
MA (Haq et al., 1987)	AGE	TROLL AREA ZONES AND SUBZONES (JGR)		Estimated boundary age (Ma)	Nearest FCSS bioevent	DIC	FCSS position	FCSS Age (Ma±2σ)	
		EARLY VOLGIAN	KIMMERIDGIAN (sensu gallico)						
140	KIMMERIDGIAN	LATE OXFORDIAN	Scriniodinium crystallinum (CR)	141	Oligosphaeridium pulcherrimum s.l. (Pu)	FDA R. cladophora	104	17.50	140.73 ± 6.2
					Leptodinium subtile (Su)	FDA S. pilosum	260	17.85	141.32 ± 7.2
					Lithodinia 'haanii' (GaH)	LDA C. granulatum	287	19.55	144.00 ± 10.8
					Glossodin. aff. dimorphum (GaD)	FDA E. galeritum	43	20.08	144.74 ± 5.8
					Endoscrinium galeritum (Ga)	FDA L. subtile (consistent)	292	20.68	145.51 ± 16.0
					Occisucysta cf. evittii (AeE)	FDA O. cf. evittii	288	21.00	145.89 ± 3.8
					Adnatosphaeridium aemulum (Ae)	FDA R. cladophora (C)	105	21.26	146.19 ± 4.0
					G. jurassica var. longicornis (AeL)	LDA Dinocyst sp. A	294	21.98	146.98 ± 2.0
					Acanthaulax senta (Se)	LDA A. cf. calloviana	291	22.04	147.05 ± 9.2
					Chytrœisphaer. cerastes (SeC)	FDA E. galeritum (C)	295	22.23	147.25 ± 5.4
145	CALLOVIAN	LOWER OXF	Systematophora divarica (SeD)	147	Lithodinia jurassica (Ju)	FDA N. pellucida	85	23.19	148.24 ± 5.4
					Acanthaulax senta (Se)	LDA Lithodinia sp. 3	266	25.05	149.83 ± 3.4
					Chytrœisphaer. cerastes (SeC)	LDA S. vestitia (consistent)	251	25.71	150.28 ± 8.6
					Systematophora divarica (SeD)	LDA G. jur. longicornis	262	26.76	150.96 ± 2.4
152	MIDDLE OXF	Acanthaulax senta (Se)	Lithodinia jurassica (Ju)	150	Lithodinia jurassica (Ju)	FDA L. scarburghensis	69	27.03	151.16 ± 2.0
					Acanthaulax senta (Se)	FDA W. fimbriata	124	27.88	151.84 ± 2.4
152	CALLOVIAN	Acanthaulax senta (Se)	Lithodinia jurassica (Ju)	151	Lithodinia jurassica (Ju)	LDA S. crystallinum	242	28.08	152.01 ± 6.4

# FCSS CORRELATION (STRATCOR)

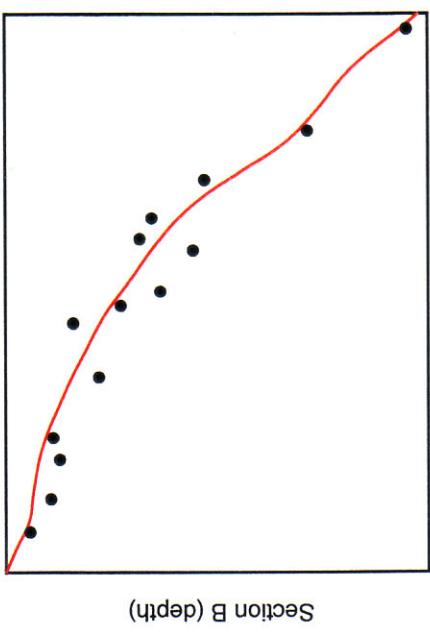
A scatter plot of events in common between the FCSS and section A is used to find A best line of correlation



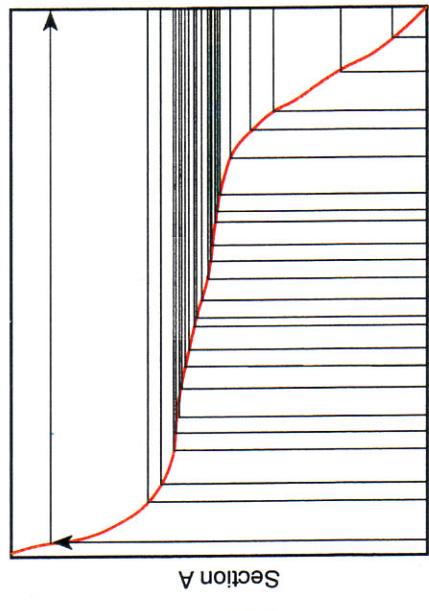
Tieing of observed bioevents often leads to ambiguities in correlations



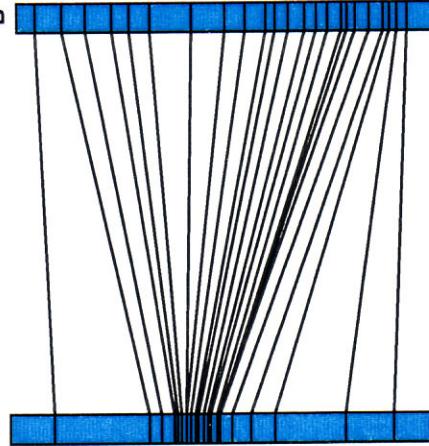
A scatter plot of events in common between the FCSS and section B is used to find B best line of correlation



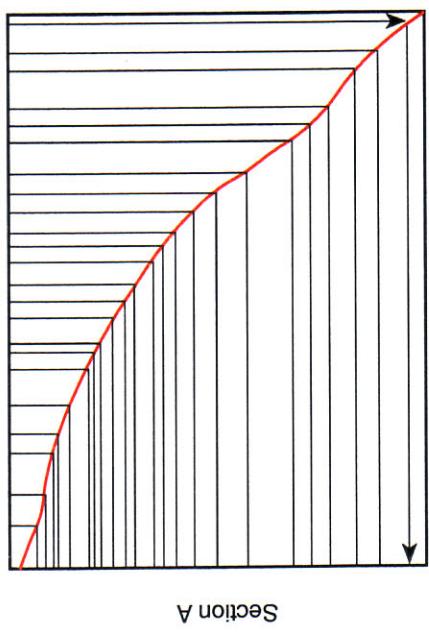
Final composite standard sequence (FCSS units or age in Ma)



High-resolution correlation is achieved by interpolating the FCSS bioevent sequence via the line of correlation for each section



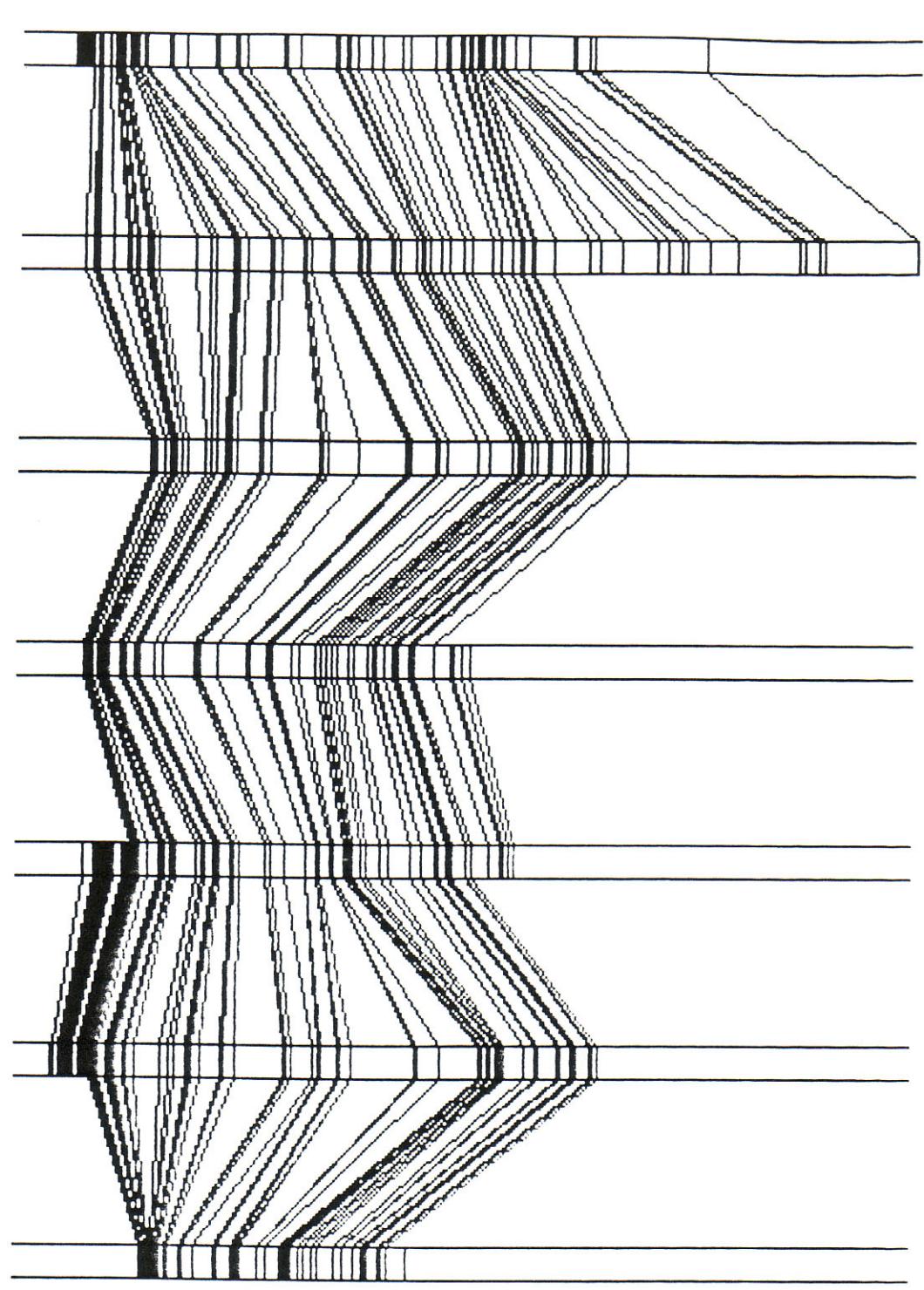
Final composite standard sequence (FCSS units or age in Ma)



Final composite standard sequence

[STRPLOT]

Scale bar  
100m



PROFILE: 5      31/2-15      31/2-9      31/2-14      31/2-11

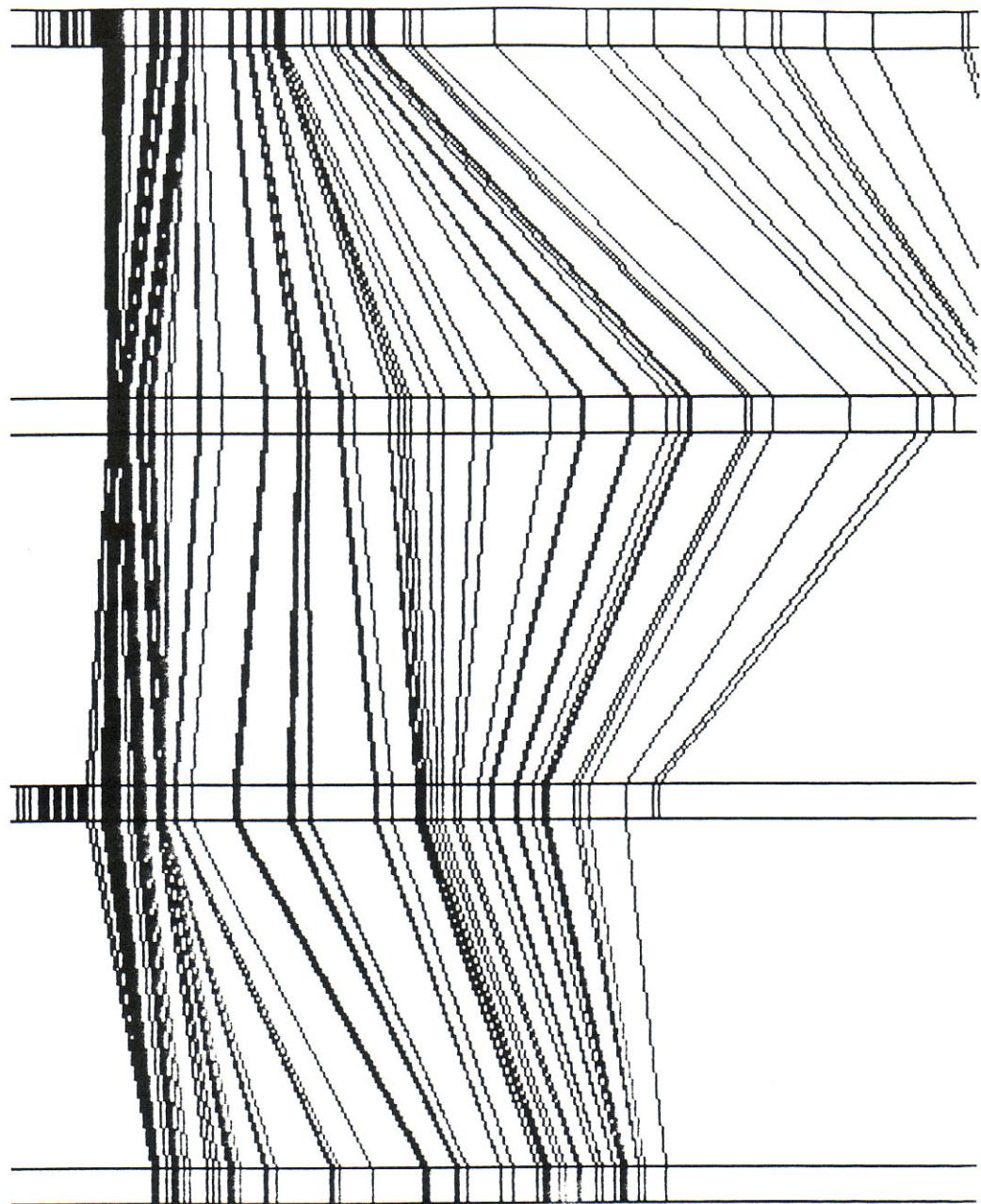
Interpolated  
bioevents

31/2-13      31/2-5      31/5-3

PROFILE: 2

Interpolated  
bioevents

31/2-13      31/2-10      31/2-3      31/3-2



Scale bar  
100m



[STRPLOT]